

Klotho Gene, Phosphocalcic Metabolism, and Survival in Dialysis

Pablo Ureña Torres,^{*,†‡} Dominique Prié,^{†‡} Laurent Beck,[‡] David De Brauwere,[‡] Christine Leroy, and Gérard Friedlander^{††}

The discovery that two recently identified molecules, klotho and fibroblast growth factor 23 (FGF23), played an important role in calcium, phosphate, and vitamin D metabolism has transformed our traditional physiological view in which bone and mineral homeostasis was mainly regulated by parathyroid hormone, vitamin D, and calcitonin, according to mineral body needs. FGF23 is a 251–amino acid secreted protein produced by osteoblasts and osteocytes in bone following the stimulation by phosphate and vitamin D or the inhibition by dentin matrix protein 1. Originally isolated from tumoral cells of patients with tumor-induced osteomalacia and hypophosphatemia, FGF23 inhibits phosphate reabsorption in renal proximal tubular cells and 1α -hydroxylase activity, resulting in decreased synthesis of calcitriol. To exert these actions, FGF23 requires the conversion, by klotho, of the canonical FGF receptor 1 (IIIc) in a specific high affinity FGF23 receptor. On the other hand, *klotho* is a putative antiaging gene identified in 1997 when a particular mouse strain, created by random insertion mutagenesis, was found to be short-lived and displayed premature atherosclerosis, osteopenia, skin atrophy, pulmonary emphysema, hyperphosphatemia, hypercalcemia, and high serum calcitriol levels. The gene of *klotho* encodes a 1012–amino acid cell-surface protein with a short cytoplasmic tail and an extracellular domain that consists in tandem duplicated copies of a β -glucuronidase-like sequence, which can be released into the circulation as soluble forms after being cleaved by metalloproteinases such as ADAM10 and ADAM17. By modulating FGF23 action, *klotho* regulates urinary phosphate excretion and calcitriol synthesis. By virtue of its β -glucuronidase activity, *klotho* deglycosylates the calcium channel TRPV5 (transient receptor potential vallinoid-5) and regulates urinary calcium excretion. *klotho* also binds to Na^+ , K^+ -ATPase in parathyroid cells and regulates calcium-stimulated PTH secretion. Finally, *klotho* extends life span via several mechanisms, including the reduction of calcitriol synthesis and serum calcium and phosphorus levels, the induction of insulin resistance, and by increasing the resistance to oxidative stress.

© 2008 by the National Kidney Foundation, Inc. All rights reserved.

The *klotho* gene was fortuitously discovered in 1997 by a Japanese group seeking antiaging genes.¹ Indeed, the *klotho* gene was disrupted in its 5′-flanking promoter region by the randomly insertion of an exogenously introduced nonfunctional gene, the rabbit Na/H exchanger under the control of the human elongation factor promoter.¹ The coding region of this mutated gene was still

preserved, but its expression was markedly reduced, generating a mouse strain with a strong hypomorphic allele. Homozygous mice for this hypomorphic allele showed shortened life span and multiple accelerated age-related disorders, including hypoactivity, sterility, skin thinning, muscle atrophy, osteoporosis, vascular calcifications, soft tissue calcifications, defective hearing, thymus atrophy, pulmonary emphysema, ataxia, and abnormalities of the pituitary gland, as well as hypoglycemia, hyperphosphatemia, and paradoxically high plasma calcitriol levels. Conversely, mice overexpressing *klotho* live 30% longer than control mice and show a slow aging process through mechanisms involving insulin resistance and increased resistance to oxidative stress.²

These animal findings have been partially confirmed in humans where several single-nucleotide polymorphisms in the *klotho* gene are associated with life span,³ osteoporosis,^{4–6} spondylosis,⁷ serum osteocalcin levels,⁸ osteonecrosis in patients with sickle cell anemia,⁹ stroke,^{3,10} and coronary

*Service de Néphrologie et Dialyse, Clinique du Landy, Saint Ouen, France.

†Service de Physiologie-Explorations Fonctionnelles, Hôpital Necker-Enfants Malades, Paris, France.

‡INSERM U845, University Paris Descartes, Centre de Recherche “Croissance et Signalisation,” Faculté de Médecine Necker, Paris, France.

Address requests for reprints to Dr. Pablo Ureña Torres, Service de Néphrologie et Dialyse, Clinique du Landy, 23 rue du Landy, 93400, Saint Ouen, France. E-mail: urena.pablo@wanadoo.fr

© 2008 by the National Kidney Foundation, Inc. All rights reserved.

1051-2276/08/0000-0000\$34.00/0

doi:10.1053/j.jrn.2008.10.018

artery diseases.^{11,12} Additionally, a homozygous missense inactivating mutation (H193R) in the *klotho* gene that was found in a 13-year-old girl has been associated with severe tumoral calcinosis with artery calcifications and marked hyperphosphatemia and hypercalcemia as well as elevated serum levels of parathyroid hormone (PTH) and fibroblast growth factor (FGF23).¹³ However, a translocation between chromosomes 9 and 13 causing increased circulating *klotho* levels results in hypophosphatemic rickets and hyperparathyroidism.¹⁴

All these observations support the assumption that *klotho* plays an important role in the selective regulation of calcium, phosphate, and active vitamin D levels, in the regulation of parathyroid mass and PTH secretion, and in aging and senescence-related disorders. This article attempts to provide an updated review of these subjects.

Molecular Characteristics of Klotho: Gene, mRNA, and Protein

The human *klotho* gene is a 5-exon gene located on chromosome 13q12 within a region longer than 50 kb.¹⁵ Two transcripts arise from this single gene: one full-length 5.2-kb encoding a 1012-amino acid (130-kDa), single-pass, membrane-anchored protein. This membrane form is released into the circulation following its cleavage at the membrane level (a-cleavage) and between the two globular glycosidase domains (b-cleavage) by several metalloproteases or sheddases such as ADAM10 and ADAM 17. The main circulating form of *klotho* is the 130- to 135-kDa, fragment which is ultimately cleaved into two smaller fragments of 65 to 68 kDa.¹⁶ The other transcript, derived from the alternative mRNA splicing, encodes the N-terminal half of *klotho*, a protein of 549 amino acids with a molecular weight of approximately 65 to 68 kDa.^{15,17,18}

Klotho protein belongs to the β -glucuronidase family. The human protein shows 86% amino acid identity with its mouse homolog. The extracellular domain of klotho is composed of two internal repeats (KL1, KL2), each of approximately 450 amino acids long with a similarity of 21% to each other. These two domains form a butterfly-shaped molecule at the surface of the cellular membrane. They share 20% to 40% sequence identity to the β -glucosidase of both bacteria and plants and with mammalian lactase glycosylceramidase.¹⁸ Another particularity of klotho proteins

is that the secreted form, as well as the membrane form, develops oligomeric complexes, suggesting a posttranslation klotho processing and possible regulatory mechanisms for klotho secretion in vivo.¹⁷

klotho mRNA is mainly expressed, in descending order, in kidney, brain, reproductive organs, pituitary gland, parathyroid glands, urinary bladder, skeletal muscle, placenta, thyroid gland, and colon.¹ In the kidney, *klotho* mRNAs and proteins are mainly localized in the distal convoluted tubule, contrasting with the major site of phosphate reabsorption and the expression of sodium-phosphate cotransporter NPT2a, which take place principally in proximal tubular cells.¹⁹ In these distal tubular cells, *klotho* colocalizes with other proteins involved in tubular calcium reabsorption such as the epithelial calcium channel TRPV5 and calbindin 28K,^{19,20} suggesting that *klotho* is intimately implicated in renal calcium homeostasis.

Mode of Action of Klotho

The exact biological role and mode of action of klotho are only partly deciphered. To date, four modes of actions have been described; first, klotho acts as a glucuronidase. This action is supported by several findings demonstrating that klotho deglycosylates synthetic and naturally occurring b-glucuronides such as b-estradiol 3b-D-glucuronide, estrone 3b-D-glucuronide, and estriol 3b-D-glucuronide,¹⁸ as well as proteins such as the calcium channels TRPV5. This enzymatic activity of klotho can be entirely mimicked by purified bovine b-glucuronidase and reduced or blocked by specific inhibitors of b-glucuronidase such as D-saccharic acid 1,4-lactone.²⁰

Second, klotho can act as a humoral factor since the secreted form and the different fragments released after cleavage of the membrane form are found in the urine, serum, and cerebrospinal fluid.²⁰ Klotho binds to a high affinity but yet not identified cell-surface klotho receptor and activates the protein kinase C (PKC) pathway in kidney and testicular cells; klotho also stimulates cAMP pathway in several cell types.²¹ The activation of this receptor by klotho leads to the inhibition of the intracellular insulin/IGF-1 signalling cascade.² This activity likely contributes to the antiaging effects of klotho, because inhibition of

insulin-like signaling is an evolutionarily conserved mechanism for extending life span.^{2,22}

Third, *klotho* can act as a coreceptor or a cofactor for other proteins such as FGF23. It has been recently demonstrated that *klotho* directly binds to multiple FGF receptors (FGFRs) and that the *klotho*/FGFR complex binds to FGF23 with higher affinity than FGFR or *klotho* alone. Furthermore, *klotho* significantly enhanced the ability of FGF23 to induce phosphorylation of FGF receptor substrate and extracellular signal-regulated kinase (ERK) in a variety of cell types.²³ The interaction between *klotho*, FGFR, and FGF23 is a new type of receptor modulation, which has been further illustrated in a recent report.²⁴ Indeed, *klotho* binds to FGFR1(IIIc) and its concerted action constitutes the FGF23 specific receptor; in the absence of *klotho*, the function of FGF23 is abolished.²⁴

Fourth, *klotho* interacts physically with Na^+, K^+ -ATPase in parathyroid cells and regulates the calcium-stimulated PTH secretion. In *klotho* knock-out mice, parathyroid Na^+, K^+ -ATPase activity is decreased, leading also to a reduced PTH secretion.²⁵ However, the molecular mechanisms for this effect of *klotho* remain to be identified.

Effects of Klotho on Bone Structure and Function

Klotho-deficient mice show low bone formation and bone resorption activities, which results in a radiographic, densitometric, and histomorphometric osteopenia.¹ The decrease in bone formation is associated with a diminution in the number of osteoblastic cells and in their ability to produce alkaline phosphatase and to mineralize extracellular matrix.²⁶ On the other hand, cultured osteoclastic cells have normal bone resorption activity and survival rate, but their differentiation process from osteoclast precursor cells is disturbed. Moreover, osteoprotegerin, a secreted factor which inhibits osteoclastogenesis, is upregulated in *klotho*-deficient mice, suggesting that there is an independent impairment of osteoblast and osteoclast differentiation, which could be the cause of this low bone turnover osteopathy.^{26,27}

In human, several single-nucleotide polymorphisms (SNPs) in the *klotho* gene have been found associated with changes in bone mineral density in Asian and Caucasian populations. In the Caucasian

population, the SNP in the promoter region (G395A) and in exon 4 (C1818T) and their haplotypes are significantly associated with low bone density in postmenopausal women (>65 years) as well as in Japanese postmenopausal women, but not in premenopausal or younger postmenopausal women. The polymorphism G395A substitution in the promoter region affects DNA-protein interaction and may affect the level of expression of *klotho*.^{4,7} Another polymorphism in *klotho* gene (F352V) has been associated with a higher bone mineral density in a Spanish population of postmenopausal women.²⁸

Effects of Klotho on Calcium, Phosphate, vitamin D, and PTH Metabolism

Mice with *klotho* gene mutation show disturbed calcium and phosphate homeostasis together with an increased in the serum concentration of active vitamin D [$1,25(\text{OH})_2\text{D}_3$]. Interestingly, most of the aging phenotypes of these mice can be lightened, as well as serum calcium and $1,25(\text{OH})_2\text{D}_3$ concentrations reduced, with phosphate restriction in the diet,^{29,30} suggesting that these phenotypes are downstream events resulting from elevated $1,25(\text{OH})_2\text{D}_3$ as shown in the FGF23 knock-out mice. Indeed, removal or reduction of $1,25(\text{OH})_2\text{D}_3$ in FGF23 and *klotho* mutant mice, by either dietary restriction or genetic manipulation, rescues premature aging-like features and ectopic calcifications.³⁰⁻³²

Calcium Metabolism

On the one hand, *klotho*-deficient mice are hypercalcemic mainly because of the hypervitaminosis D and its stimulatory effects on intestinal and renal absorption of calcium. However, concomitantly with the rise in serum calcium concentration, these animals exhibit an increase in urinary fractional excretion of calcium, despite low serum PTH levels, which suggests that the hypercalcemia is not entirely due to abnormal renal calcium handling. Another possible explanation is that the low bone remodeling observed with the lack of *klotho* could favor the occurrence of hypercalcemia because the skeleton would be unable to provide its buffering action. Moreover, it has been demonstrated that TRPV5 is essential for a proper osteoclastic activity; mice lacking TRPV5 have an increase in osteoclast size and number, but bone

resorption is diminished because of a reduced activity.³³ Klotho could also exert a similar effect on osteoclast TRPV5 expression as in the kidney cells but that remains to be investigated.

On the other hand, there are evidence that klotho directly participates in the regulation of renal calcium reabsorption. Klotho regulates calcium reabsorption in the distal convoluted tubule via a novel molecular mechanism, by deglycosylating and stabilizing the epithelial calcium channel TRPV5 at the cellular membrane surface.²⁰ Klotho colocalizes with TRPV5 and calbindin-D28K in the distal convoluted and connecting tubule of mouse kidney cells, which are nephron segments responsible for active transepithelial calcium reabsorption.²⁰ However, the lack of klotho leads to a diminution in the expression of TRPV5 at the cell surface and reduced tubular calcium reabsorption. Likewise, mice lacking TRPV5 have reduced klotho expression and diminished renal calcium reabsorption despite enhanced levels of $1,25(\text{OH})_2\text{D}_3$. Although the two proteins together with calbindin-D28K are tightly controlled by vitamin D, suggesting a functional link between these proteins in the maintenance of calcium homeostasis, the renal origin of the hypercalcemia of the klotho null mouse appears to be more unlikely.^{20,30}

Phosphate Metabolism

Hyperphosphatemia observed in *klotho* mutant mice is also principally due to hypervitaminosis D. However, other mechanisms could also be involved because of the major role played by klotho in phosphate metabolism. In this regard, *klotho* mutant mice display increased expression and activity of intestinal sodium-dependent phosphate cotransporter type IIb (NPT2b) and increased intestinal phosphate absorption compared with wild-type mice.³⁴ *klotho* mutant mice also have increased activity of renal sodium-dependent phosphate cotransporters NPT2a and NPT2c compared with wild-type mice, which corroborates the role played by the kidney, with the increased tubular phosphate reabsorption, in hyperphosphatemia.³⁴ Moreover, a low phosphate diet results in an increase in renal NPT2a expression in normal mice but not in *klotho* mice, suggesting that a deregulation of expression and trafficking of NPT2a play a major role in this hyperphosphatemia.³⁴ Klotho mice have serum

FGF23 levels 150- to 2,000fold higher than wild-type animals, and a low phosphate diet decreases FGF23 levels, which suggests that FGF23 cannot exert its phosphaturic effect correctly in the absence of klotho.^{24,34} The mechanism behind this phenomenon has now been elucidated; klotho binds to FGFR1(IIIc) and its concerted action constitutes the FGF23-specific receptor. Therefore, klotho functions as a cofactor essential for the stimulation of FGFR1(IIIc) by FGF23 and in this way modulates the phosphaturic effect of FGF23.²⁴

Vitamin D Metabolism

Klotho-deficient mice show low serum $24,25(\text{OH})_2\text{D}_3$ and increased $1,25(\text{OH})_2\text{D}_3$ concentrations, which are due to an increase in both the renal 25-hydroxyvitamin D-1 α -hydroxylase (CYP27b1) and the 24-hydroxylase activities. Of note, in these animals, the normal pathways leading to the upregulation of CYP27b1, such as PTH, calcitonin, and $1,25(\text{OH})_2\text{D}_3$, are intact, suggesting the existence of other regulatory pathways.³⁰ Like $1,25(\text{OH})_2\text{D}_3$, dietary phosphate depletion, a recognized stimulus of CYP27b1 expression, also increases the renal expression of klotho; supporting again the hypothesis that klotho could influence renal CYP27b1 expression.

PTH Metabolism

PTH is produced by the parathyroid glands in response to low extracellular ionized calcium concentration and through the stimulation of the G protein-coupled calcium-sensing receptor.³⁵ However, the molecular mechanisms behind the regulation of PTH secretion by low calcium concentration are not yet fully elucidated and it has been suggested that Na^+, K^+ -ATPase could be one of them. Na^+, K^+ -ATPase is essential in maintaining intracellular Na^+ and K^+ concentrations and in creating the transmembrane concentration gradient for the transport across the membrane of many substrates. A high Na^+ gradient created by the Na^+, K^+ -ATPase activity has been proposed to drive the transepithelial calcium transport in the choroid plexus and the kidney, and probably now in the parathyroid gland. A recent study has demonstrated that klotho interacts physically with Na^+, K^+ -ATPase in parathyroid cells and regulates calcium-stimulated PTH secretion.²⁵ In addition, *klotho* mutant mice show a reduction of

Q7

27% in the PTH secretion in response to induced hypocalcemia, and the production of PTH by isolated parathyroid cells in a low calcium medium is decreased, similar to the reduction observed in normal parathyroid cells treated by ouabain, a specific inhibitor of Na^+, K^+ -ATPase. However, the addition of ouabain to parathyroid cells from the *klotho* mutant mice does not further decrease PTH production. These data suggest that *klotho* may play an important role in the Na^+, K^+ -ATPase-dependent release of PTH; however, the molecular mechanisms for this role of *klotho* remain to be further investigated.

Klotho may also regulate parathyroid glandular mass and function. This is suggested by the recent observation of elevated circulating *klotho* levels, hypophosphatemic rickets, and hyperparathyroidism in a young girl with a genomic translocation between chromosomes 9 and 13.¹⁴ Interestingly, the increase in *klotho* level appeared to be sufficient to produce hyperparathyroidism and also PTH-independent hypophosphatemia. As circulating levels of FGF23 were markedly increased in this patient, and given that *klotho* serves as a cofactor for FGF23, that FGF23 inhibits the expression of PTH mRNA and secretion of PTH in parathyroid cells,³⁶ and that *klotho* regulates PTH secretion through the maintenance of cell surface Na^+, K^+ -ATPase activity,²⁵ it can be suggested the existence of a very complex interaction between these molecules in the regulation of PTH metabolism.

Klotho and Longevity

Klotho-deficient mice exhibit a syndrome resembling human premature aging, with multiple pathological phenotypes in tissues including reproductive organs. This phenotype can be rescued by exogenous expression of *klotho* cDNA¹ and, interestingly, *klotho* gene overexpression extends life span by 20% to 30%.² In humans, in a population-based association study, using two microsatellite markers flanking the *klotho* gene and DNA sequencing, it was revealed that a functional variant of *klotho* (KL-VS) was associated with human survival, defined as postnatal life expectancy (>75 years) and longevity.³⁷ In addition, there is a progressive decline with aging of serum *klotho* levels, as assessed by a recent ELISA using a polyclonal antibody against the C-terminal of human secreted *klotho* protein.³⁸

Klotho extends life span by inhibiting the aging process, probably through several mechanisms, including the induction of insulin resistance.² Indeed, by inducing the inhibition of insulin/IGF-1 signaling, *klotho* also increases the resistance to oxidative stress at the cellular and subcellular level in mammals. Furthermore, *klotho* protein activates the FoxO forkhead transcription factors that are negatively regulated by insulin/IGF-1 signaling, thereby inducing expression of manganese superoxide dismutase. This in turn facilitates removal of reactive oxygen species and confers oxidative stress resistance.^{22,39}

Klotho could also extend life span by protecting the cardiovascular system through endothelium-derived NO production. Many experimental data support this hypothesis: *klotho* reduces H_2O_2 -induced apoptosis and cellular senescence in vascular cells⁴⁰ and the impaired endothelium-dependent vasodilation of the aorta and arterioles of heterozygous *klotho*-deficient mice are restored by parabiosis with wild-type mice⁴¹ or by in vivo *klotho* gene delivery.^{42,43} The *klotho*-induced insulin resistance could prevent cellular lipid overload by reducing insulin-stimulated availability of the lipogenic substrate glucose and thereby could decrease the cellular apoptosis.⁴⁴

Finally, the extended life span observed in animal overexpressing *klotho* may also be due to the reduction of calcitriol synthesis, serum calcium, and phosphorus levels. This is supported by the rescue of the phenotype by removal or reduction of $1,25\text{OH}_2\text{D}_3$ in FGF23 and *klotho* mutant mice, by either dietary restriction or genetic manipulation.^{30,32}

Conclusion

The discovery of *klotho*, which is indeed a newly recognized hormone, has opened an extraordinary field of investigation not only because of its implications in human longevity but also because of its implication in a multitude of other biological processes. Although the exact biological role and molecular function of *klotho* have been only partly deciphered, several functions seem to be clearly established. *Klotho* is an antiaging molecule, which binds to a not yet identified high affinity receptor and inhibits the intracellular insulin/IGF-1 signaling cascade; *klotho* functions as a β -glucuronidase that deglycosylates

steroid β -glucuronides as well as the calcium channels TRPV5; klotho is a cofactor essential for the activation of FGFR1(IIIc) signalling by FGF23. Circulating klotho plays certainly an important role in a variety of metabolic syndromes including in patients with chronic kidney disease and hyperphosphatemia.

References

1. Kuro-o M, Matsumura Y, Aizawa H, et al: Mutation of the mouse klotho gene leads to a syndrome resembling ageing. *Nature* 390:45-51, 1997
2. Kurosuo H, Yamamoto M, Clark JD, et al: Suppression of aging in mice by the hormone Klotho. *Science* 309:1829-1833, 2005
3. Arking DE, Atzmon G, Arking A, et al: Association between a functional variant of the KLOTHO gene and high-density lipoprotein cholesterol, blood pressure, stroke, and longevity. *Circ Res* 96:412-418, 2005
4. Kawano K, Ogata N, Chiano M, et al: Klotho gene polymorphisms associated with bone density of aged postmenopausal women. *J Bone Miner Res* 17:1744-1751, 2002
5. Kawano K, Kawaguchi H: Human klotho gene polymorphisms associated with bone density of aged postmenopausal women *Nippon Rinsho* 62 Suppl 2:190-194, 2004
6. Yamada Y, Ando F, Niino N, et al: Association of polymorphisms of the androgen receptor and klotho genes with bone mineral density in Japanese women. *J Mol Med* 83:50-57, 2005
7. Ogata N, Matsumura Y, Shiraki M, et al: Association of klotho gene polymorphism with bone density and spondylosis of the lumbar spine in postmenopausal women. *Bone* 31:37-42, 2002
8. Mullin BH, Wilson SG, Islam FM, et al: Klotho gene polymorphisms are associated with osteocalcin levels but not bone density of aged postmenopausal women. *Calcif Tissue Int* 77:145-151, 2005
9. Baldwin C, Nolan VG, Wyszynski DE, et al: Association of klotho, bone morphogenic protein 6, and annexin A2 polymorphisms with sickle cell osteonecrosis. *Blood* 106:372-375, 2005
10. Kim Y, Kim JH, Nam YJ, et al: Klotho is a genetic risk factor for ischemic stroke caused by cardioembolism in Korean females. *Neurosci Lett* 407:189-194, 2006
11. Arking DE, Becker DM, Yanek LR, et al: KLOTHO allele status and the risk of early-onset occult coronary artery disease. *Am J Hum Genet* 72:1154-1161, 2003
12. Imamura A, Okumura K, Ogawa Y, et al: Klotho gene polymorphism may be a genetic risk factor for atherosclerotic coronary artery disease but not for vasospastic angina in Japanese. *Clin Chim Acta* 371:66-70, 2006
13. Ichikawa S, Imel EA, Kreiter ML, et al: A homozygous missense mutation in human KLOTHO causes severe tumoral calcinosis. *J Musculoskelet Neuronal Interact* 7:318-319, 2007
14. Brownstein CA, Adler F, Nelson-Williams C, et al: A translocation causing increased alpha-klotho level results in hypophosphatemic rickets and hyperparathyroidism. *Proc Natl Acad Sci U S A* 105:3455-3460, 2008
15. Matsumura Y, Aizawa H, Shiraki-Iida T, et al: Identification of the human klotho gene and its two transcripts encoding membrane and secreted klotho protein. *Biochem Biophys Res Commun* 242:626-630, 1998
16. Chen CD, Podvin S, Gillespie E, et al: Insulin stimulates the cleavage and release of the extracellular domain of Klotho by ADAM10 and ADAM17. *Proc Natl Acad Sci U S A* 104:19796-19801, 2007
17. Kato Y, Arakawa E, Kinoshita S, et al: Establishment of the anti-Klotho monoclonal antibodies and detection of Klotho protein in kidneys. *Biochem Biophys Res Commun* 267:597-602, 2000
18. Tohyama O, Imura A, Iwano A, et al: Klotho is a novel beta-glucuronidase capable of hydrolyzing steroid beta-glucuronides. *J Biol Chem* 279:9777-9784, 2004
19. Li SA, Watanabe M, Yamada H, et al: Immunohistochemical localization of Klotho protein in brain, kidney, and reproductive organs of mice. *Cell Struct Funct* 29:91-99, 2004
20. Chang Q, Hoefs S, van der Kemp AW, et al: The beta-glucuronidase klotho hydrolyzes and activates the TRPV5 channel. *Science* 310:490-493, 2005
21. Imai M, Ishikawa K, Matsukawa N, et al: Klotho protein activates the PKC pathway in the kidney and testis and suppresses 25-hydroxyvitamin D3 1alpha-hydroxylase gene expression. *Endocrine* 25:229-234, 2004
22. Yamamoto M, Clark JD, Pastor JV, et al: Regulation of oxidative stress by the anti-aging hormone klotho. *J Biol Chem* 280:38029-38034, 2005
23. Kurosuo H, Ogawa Y, Miyoshi M, et al: Regulation of fibroblast growth factor-23 signaling by klotho. *J Biol Chem* 281:6120-6123, 2006
24. Urakawa I, Yamazaki Y, Shimada T, et al: Klotho converts canonical FGF receptor into a specific receptor for FGF23. *Nature* 444:770-774, 2006
25. Imura A, Tsuji Y, Murata M, et al: alpha-Klotho as a regulator of calcium homeostasis. *Science* 316:1615-1618, 2007
26. Kawaguchi H, Manabe N, Miyaura C, et al: Independent impairment of osteoblast and osteoclast differentiation in klotho mouse exhibiting low-turnover osteopenia. *J Clin Invest* 104:229-237, 1999
27. Kawaguchi H, Manabe N, Chikuda H, et al: Cellular and molecular mechanism of low-turnover osteopenia in the klotho-deficient mouse. *Cell Mol Life Sci* 57:731-737, 2000
28. Riancho JA, Valero C, Hernandez JL, et al: Association of the F352V variant of the Klotho gene with bone mineral density. *Biogerontology* 8:121-127, 2007
29. Morishita K, Shirai A, Kubota M, et al: The progression of aging in klotho mutant mice can be modified by dietary phosphorus and zinc. *J Nutr* 131:3182-3188, 2001
30. Tsujikawa H, Kurotaki Y, Fujimori T, et al: Klotho, a gene related to a syndrome resembling human premature aging, functions in a negative regulatory circuit of vitamin D endocrine system. *Mol Endocrinol* 17:2393-2403, 2003
31. Razzaque MS, Sitara D, Taguchi T, et al: Premature aging-like phenotype in fibroblast growth factor 23 null mice is a vitamin D-mediated process. *Faseb J* 20:720-722, 2006
32. Razzaque MS, Lanske B: Hypervitaminosis D and premature aging: lessons learned from Fgf23 and Klotho mutant mice. *Trends Mol Med* 12:298-305, 2006
33. van der Eerden BC, Hoenderop JG, et al: The epithelial Ca2+ channel TRPV5 is essential for proper osteoclastic bone resorption. *Proc Natl Acad Sci U S A* 102:17507-17512, 2005
34. Segawa H, Yamanaka S, Ohno Y, et al: Correlation between hyperphosphatemia and type II Na-Pi cotransporter activity in klotho mice. *Am J Physiol Renal Physiol* 292:F769-779, 2007

35. Brown E, Gamba G, Riccardi D, et al: Cloning and characterization of an extracellular Ca^{2+} -sensing receptor from bovine parathyroid. *Nature* 366:575-580, 1993
36. Krajisnik T, Bjorklund P, Marsell R, et al: Fibroblast growth factor-23 regulates parathyroid hormone and 1 α -hydroxylase expression in cultured bovine parathyroid cells. *J Endocrinol* 195: 125-131, 2007
37. Arking DE, Krebsova A, M. Macek Sr, et al: Association of human aging with a functional variant of klotho. *Proc Natl Acad Sci U S A* 99:856-861, 2002
38. Xiao NM, Zhang YM, Zheng Q, et al: Klotho is a serum factor related to human aging. *Chin Med J (Engl)* 117:742-747, 2004
39. Kappeler L, De Magalhaes Filho C, et al: Ageing, genetics and the somatotrophic axis *Med Sci (Paris)* 22:259-265, 2006
40. Ikushima M, Rakugi H, Ishikawa K, et al: Anti-apoptotic and anti-senescence effects of Klotho on vascular endothelial cells. *Biochem Biophys Res Commun* 339:827-832, 2006
41. Nagai R, Saito Y, Ohyama Y, et al: Endothelial dysfunction in the klotho mouse and downregulation of klotho gene expression in various animal models of vascular and metabolic diseases. *Cell Mol Life Sci* 57:738-746, 2000
42. Saito Y, Yamagishi T, Nakamura T, et al: Klotho protein protects against endothelial dysfunction. *Biochem Biophys Res Commun* 248:324-329, 1998
43. Saito Y, Nakamura T, Ohyama Y, et al: In vivo klotho gene delivery protects against endothelial dysfunction in multiple risk factor syndrome. *Biochem Biophys Res Commun* 276:767-772, 2000
44. Unger RH: Klotho-induced insulin resistance: a blessing in disguise? *Nat Med* 12:56-57, 2006